



King's Research Portal

DOI:

[10.1007/s00431-018-3090-5](https://doi.org/10.1007/s00431-018-3090-5)

Document Version

Peer reviewed version

[Link to publication record in King's Research Portal](#)

Citation for published version (APA):

Dassios, T., Kaltsogianni, O., & Hickey, A. (2018). Deltoid muscle morphometry as an index of impaired skeletal muscularity in neonatal intensive care. *European Journal of Pediatrics*. <https://doi.org/10.1007/s00431-018-3090-5>

Citing this paper

Please note that where the full-text provided on King's Research Portal is the Author Accepted Manuscript or Post-Print version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version for pagination, volume/issue, and date of publication details. And where the final published version is provided on the Research Portal, if citing you are again advised to check the publisher's website for any subsequent corrections.

General rights

Copyright and moral rights for the publications made accessible in the Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognize and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Research Portal

Take down policy

If you believe that this document breaches copyright please contact librarypure@kcl.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

**Deltoid muscle morphometry as an index of impaired skeletal muscularity in neonatal
intensive care**

**Theodore Dassios^{1,2,3}, Ourania Kaltsogianni¹, Miltiadis Krokidis⁴,
Ann Hickey¹, Anne Greenough^{2,3,5}**

Email address of authors:

Theodore.dassios@nhs.net

Raniagk1@hotmail.com

Miltiadis.krokidis@addenbrookes.nhs

annhickey@nhs.net

anne.greenough@kcl.ac.uk

Address for Correspondence: Professor Anne Greenough, NICU, 4th Floor Golden
Jubilee Wing, King's College Hospital, Denmark Hill, London, SE5 9RS, Tel: 0203 299
3037; fax: 0203 299 8284; email: anne.greenough@kcl.ac.uk

¹ Neonatal Intensive Care Centre, King's College Hospital NHS Foundation Trust, London, UK;

² MRC-Asthma UK Centre in Allergic Mechanisms of Asthma, King's College London, UK;

³ Women and Children's Health, School of Life Course Sciences, Faculty of Life Sciences and Medicine, King's
College London, UK;

⁴ Department of Radiology, Cambridge University Hospitals NHS Foundation Trust, Cambridge, UK;

⁵ National Institute for Health Research (NIHR) Biomedical Research Centre based at Guy's and St Thomas' NHS
Foundation Trust and King's College London; UK

ABSTRACT

We hypothesised that extremely premature infants would have decreased muscle mass at term-corrected age compared to term-born infants and that the degree of reduced muscle mass acquisition would correlate with the duration of invasive mechanical ventilation. The MRI brain scans of infants admitted in the neonatal unit at King's College Hospital between 1/1/2010 and 1/6/2016 were retrospectively reviewed. The coronal cross-sectional area of the left deltoid muscle (DCSA) was measured in 17 infants born <28 weeks of gestation and in 20 infants born at term. The prematurely born infants had a median(IQR) gestation age of 25(24-27) weeks and the term infants 40(38-41) weeks. The duration of invasive mechanical ventilation for the prematurely born infants was 39(14-62) days and for the term infants for 4(2-5) days, $p<0.001$. DCSA was smaller in prematurely-born infants (median 189, IQR:176-223mm²) compared to term-born infants (median 302, IQR 236-389mm²), $p<0.001$. DCSA was related to gestation age ($r=0.545$, $p=0.001$), weight z-score at MRI ($r=0.658$, $p<0.001$) and days of invasive mechanical ventilation ($r=-0.583$, $p<0.001$). In conclusion, extremely premature infants studied at term had a lower muscle mass compared to term-born infants.

Conclusion: Our results suggest prolonged mechanical ventilation in infants admitted in neonatal intensive care is associated with reduced skeletal muscle mass acquisition.

KEY WORDS: Deltoid muscle morphometry, impaired skeletal muscularity, neonatal intensive care

49 **ABBREVIATIONS**

50 DCSA Deltoid cross-sectional area

51 IQR Interquartile range

52 MRI Magnetic Resonance Imaging

53

54 **AUTHORS SUMMARY**

55

56 **What is known:**

- 57 • Prolonged mechanical ventilation in adult intensive care patients has been
58 associated with skeletal muscle dysfunction and atrophy.
- 59 • The cross-sectional area of the deltoid muscle has been used to evaluate muscle
60 atrophy in infants with a previous brachial plexus birth injury.

61

62

63 **What is new:**

- 64 • Premature infants studied at term exhibit lower cross-sectional area of the deltoid
65 muscle than their term counterparts.
- 66 • Prolonged mechanical ventilation could be associated with skeletal muscle
67 impairment.

68

INTRODUCTION

Following birth, prematurely-born infants commonly spend a long period in neonatal intensive care where they often undergo invasive mechanical ventilation. The effect of critical illness on the skeletal muscles, commonly referred to as “critical illness myopathy”, has been well-described in adults. The most common predisposing conditions in adult intensive care patients are acute respiratory disorders such as acute respiratory distress syndrome, pneumonia and severe asthma [6, 13]. In addition, use of high-dose steroids, non-depolarizing blocking agents [2], aminoglycosides [19] and acidosis [14] may also predispose to critical illness myopathy.

In prematurely born neonates, the postnatal nutritional target is the accretion of body weight at a rate similar to that achieved during intrauterine development [1]; that gold standard, however, is infrequently achieved [4, 5] especially in the sickest infants who rely for long periods on parenteral nutrition. The “weight-targeted” nutritional approach fails to detect qualitative differences in body composition and weight accretion, predominantly in the form of adipose tissue, does not necessarily translate to better outcomes [9]. This is particularly relevant in the prematurely born population whose adult-life adiposity profile might place them at an increased risk for cardiovascular and metabolic disease [15].

Few studies have reported data on qualitative body composition in prematurely born compared to term-born infants [16] and to our knowledge none has specifically investigated skeletal muscle mass. The magnetic resonance imaging (MRI) appearance of the deltoid muscle has been used to evaluate muscle atrophy in infants with a previous branchial plexus birth injury [10, 17]. The cross-sectional area of the deltoid muscle (DCSA) on the affected

side has been shown to be significantly decreased compared to the unaffected size with a mean affected to normal ratio of 76% [17]. The deltoid muscle is often included in the coronal images of MRI-brain scans that prematurely born infants have to assess ventricular dilatation and periventricular leukomalacia and term infants for evidence of white matter injury secondary to hypoxic ischemic encephalopathy (HIE).

We hypothesised that DCSA would be smaller in prematurely born infants studied at term compared to term born infants and that the DCSA would correlate with body weight and gestational age. We also hypothesised that DCSA would correlate with days of ventilation and days of parenteral nutrition. Our aims were to test those hypotheses.

METHODOLOGY

Subjects

The MRI brain scans of infants that were admitted to the neonatal unit at King's College Hospital NHS Foundation Trust, London, UK and underwent a scan between 1 January 2010 and 1 June 2016 were retrospectively reviewed. Two groups of infants were assessed: term-born infants with HIE and extremely prematurely born infants (<28 weeks of gestation at birth) scanned at term that had periventricular leukomalacia or ventricular dilatation diagnosed and sequentially assessed by cranial ultrasonography. The Research and Development department confirmed that the data collection was consistent with a service evaluation/audit, and as such did not require research ethics approval. The study was registered with the Clinical Governance Department of King's College Hospital NHS Foundation Trust.

Calculation of DCSA

Infants were scanned on a 1.5 T MR imager (Siemens, Erlanger, Germany). After localizer sequences, T1-weighted in axial, oblique coronal (angled to the temporal lobe) and oblique sagittal planes were obtained (TR/TE 4000/126 msec: base resolution: 256 mm). T2-weighted images in axial, oblique coronal (angled to the temporal lobe) were obtained (TR/TE 4000/126 msec: base resolution: 256 mm. The slice thickness was 4 mm for all sequences. The field of view was 200x200 mm for T1-weighted images and 220x220 mm for T2-weighted images).

The deltoid muscle was visualised on the oblique coronal plane at the level where the head of the humerus had the maximum diameter at which level the area of the deltoid is maximal [17]. Free-hand tracing of the perimeter of the left deltoid muscle was undertaken by a consultant radiologist (MK) and the DCSA was calculated in mm² by the Sectra PACS software (Sectra AB, Linköping, Sweden) (Figure 1). The radiologist performing the DCSA measurement was unaware of the infant's clinical condition and demographics.

Information from the medical records

Gender, gestation age at birth, birth weight and postmenstrual age, postnatal age and weight at the time of MRI were recorded. Data also recorded were whether the infant had had postnatal steroids or confirmed necrotising enterocolitis, the days of invasive mechanical ventilation and parenteral nutrition prior to the MRI and the use of non-depolarising muscle relaxing agents.

Statistical analysis

Data were tested for normality with the Kolmogorov–Smirnov test and found to be non-normally distributed. Continuous data are presented as median and interquartile range (IQR). Differences between term and premature infants were tested for significance using the Mann-Witney rank sum test. Multiple linear regression was used to adjust for confounders in differences of DCSA between term and prematurely born infants. Variables without normal distribution were logarithmically transformed. Multi-collinearity among the independent variables in the multiple regression analysis was assessed by calculation of the tolerance for the independent variables. Kendall's tau rank correlation coefficient (τ) was used to examine the relationship of DCSA with gestational age, age at MRI, birth weight, weight at measurement, days of ventilation and days of parenteral nutrition. Multiple regression analysis was used to examine the independent effect on DCSA of the parameters that were significantly related to DCSA in the bivariate analysis. Parameters with a variance inflation factor (VIF) of more than three were considered collinear.

Linear regression analysis was used to examine the relationship of DCSA to weight at MRI. The type of non-linearity was tested by visual inspection of the residuals. Statistical analysis was performed using IBM SPSS Software (IBM, Chicago IL).

RESULTS

Between 1 January 2010 and 1 June 2016, 291 infants were admitted to the neonatal unit with a gestational age of less than 28 weeks; 17 had an MRI brain scan to assess the extent of periventricular leukomalacia or ventricular dilatation. Twenty-one term infants underwent an MRI brain scan for HIE. One term infant was excluded from the study because the coronal views did not fully include the left deltoid muscle. Premature infants did not differ

from term infants in terms of gender, birth weight z-score, weight at MRI and administration of postnatal steroids. Prematurely born infants had longer durations of mechanical ventilation and parenteral nutrition and lower weight z-score at MRI than the term infants (table 1). Premature infants had lower DCSA compared to term infants (table 1) and lower DCSA compared to term infants after adjusting for weight at MRI ($p=0.021$, 95% Confidence Intervals: 9.628 – 111.843, Odds ratio: 0.343). DCSA was significantly related to gestational age ($\tau=0.545$, $p=0.001$), birth weight z-score ($\tau=0.547$, $p=0.001$), age at MRI ($\tau=-0.590$, $p<0.001$), weight z-score at MRI ($\tau=0.658$, $p<0.001$), days of mechanical ventilation ($\tau=-0.583$, $p<0.001$), and days of parenteral nutrition ($\tau=-0.617$, $p<0.001$). Following multiple regression analysis with DCSA as the outcome variable, corrected GA was excluded from the model due to collinearity with GA (VIF: 3.29) and days of parenteral nutrition were excluded from the model due to collinearity with days of mechanical ventilation (VIF: 4.32). DCSA remained significantly related to GA (adjusted $p=0.003$) and days of mechanical ventilation (adjusted $p=0.008$).

The linear regression analysis of DCSA against weight z-score at MRI is presented in figure 2. The quadratic model was found to accommodate the best non-linear fit (r^2 quadratic = 0.576).

DISCUSSION

We have demonstrated that DCSA is smaller in prematurely-born infants studied at term compared to term-born infants and that the DCSA is related to gestation, weight, duration of mechanical ventilation and duration of parenteral nutrition. Our results are in agreement with previous studies that have described that respiratory muscle strength in prematurely-born infants increases with increasing maturity [3]. Furthermore, the deleterious effect of prolonged mechanical ventilation on skeletal muscle function has been reported in adult

intensive care studies [6, 13]. In this paper we further highlight that prematurity, prolonged mechanical ventilation and parenteral nutrition (PN) are associated with reduced skeletal muscle mass acquisition. The importance of our study is that it is the first study to highlight that extremely premature infants exhibit postnatal skeletal muscle growth impairment which is related to prolonged mechanical ventilation. The duration of PN might directly represent a nutritional deficit or it might indirectly describe the cumulative severity of the intensive care stay as sicker infants will fail to advance on enteral nutrition and will require longer courses of PN.

Muscle atrophy and prolonged mechanical ventilation have an adverse synergistic relationship. Prolonged mechanical ventilation is associated with a release of pro-inflammatory cytokines [11] and the use of non-depolarizing muscle relaxants [2]. Inversely, sepsis [12] and disuse atrophy [18] directly impact on respiratory muscle function and lead to inability to wean off respiratory support. A plausible pathophysiological mechanism that explains our findings is that prolonged intensive care leads to a generalised skeletal myopathy with atrophy and necrosis of the muscle fibres and loss of myelinated nerve fibres [7, 8].

One implication of our study is that weight accretion alone might not be an adequate nutritional target in neonatal intensive care and that other anthropometric parameters should be taken into account in evaluating postnatal growth. These could include body composition analysis either by dual-energy X-ray absorptiometry, whole body MRI or bioelectrical impedance analysis. Those methods, however, have their own disadvantages such as high cost, non-availability and technical difficulties of performing serial measurements. The DCSA was indeed significantly related to weight at MRI however, given that they are both anthropometric indices, one might expect a stronger correlation. This relatively weak correlation would support the conclusion that body weight might not be the most appropriate

variable to reflect the nutritional status of these infants. Possibly, indices that incorporate information on height such as the ponderal index or the body mass index might be more appropriate.

The strengths of our study include that the radiologist who performed the DCSA measurements was blinded to the demographic and clinical details and hence was not biased in estimating the DCSA. We should acknowledge as a limitation the retrospective nature of our study and that we evaluated a selected subpopulation of premature infants that had significant intracranial pathology detected on cranial ultrasound. As such, these infants might have had a relatively more severe intensive care course. It is plausible that equally premature infants without intracranial pathology might have exhibited less prominent muscle atrophy. Our positive results emphasize the interest to study a "well" preterm population to demonstrate the independent impact of prolonged mechanical ventilation on these infants. We should also note as a limitation that the radiologist that evaluated the MRIs had access to the full images including the brain appearances. We should also note as a limitation that the radiologist that evaluated the MRIs had access to the full images including the brain appearances. The appearances of PVL or ventricular dilatation, therefore, might have had an impact on the blinding process. We thus suggest that future studies could eliminate this bias by excluding the brain from the reviewed images.

In conclusion, we have demonstrated that extremely premature infants do not accumulate/accrete muscle mass at a rate similar to their term counterparts and that prolonged ventilation in these infants is related to reduced skeletal muscle mass acquisition.

ACKNOWLEDGEMENTS

Compliance with ethical standards

Funding: The research was supported by the National Institute for Health Research (NIHR) Biomedical Research Centre based at Guy's and St Thomas' NHS Foundation Trust and King's College London. The views expressed are those of the author(s) and not necessarily those of the NHS, the NIHR or the Department of Health.

Compliance and ethical standards: The Research and Development department confirmed that the data collection was consistent with a service evaluation/audit, and as such did not require research ethics approval. The study was registered with the Clinical Governance Department of King's College Hospital NHS Foundation Trust.

Conflict of interest: The authors have no financial relationship relevant to this article to disclose.

Informed consent: Not required.

Contributors' statement:

T.D. conceived the study, participated in the analysis of the data, and drafted the first version of the article.

O.K. collected the data and participated in the analysis of the data.

M.K. independently reviewed the MRI scans and critically reviewed the manuscript.

A.H. contributed to study design, writing of the manuscript and interpretation of the results.

264 A.G. supervised the project, contributed to the study design and interpretation of the results,
265 and critically revised the manuscript.

266 All authors were involved in the preparation of the manuscript and approved the final
267 manuscript as submitted.

268

269

REFERENCES

1. Agostoni C, Buonocore G, Carnielli VP, De Curtis M, Darmaun D, Decsi T, Domellöf M, Embleton ND, Fusch C, Genzel-Boroviczeny O, Goulet O, Kalhan SC, Kolacek S, Koletzko B, Lapillonne A, Mihatsch W, Moreno L, Neu J, Poindexter B, Puntis J, Putet G, Rigo J, Riskin A, Salle B, Sauer P, Shamir R, Szajewska H, Thureen P, Turck D, van Goudoever JB, Ziegler EE; ESPGHAN Committee on Nutrition (2010) Enteral nutrient supply for preterm infants: commentary from the European Society of Paediatric Gastroenterology, Hepatology and Nutrition Committee on Nutrition. *J Pediatr Gastroenterol Nutr* 50:85-91.
2. Barohn RJ, Jackson CE, Rogers SJ, Ridings LW, McVey AL (1994) Prolonged paralysis due to nondepolarizing neuromuscular blocking agents and corticosteroids. *Muscle Nerve* 17:647-654.
3. Dimitriou G, Greenough A, Rafferty GF, Moxham J (2001) Effect of maturity on maximal transdiaphragmatic pressure in infants during crying. *Am J Respir Crit Care Med* 164:433-436.
4. Dusick AM, Poindexter BB, Ehrenkranz RA, Lemons JA (2003) Growth failure in the preterm infant: can we catch up? *Sem Perinatol* 27:302-310.
5. Griffin IJ, Tancredi DJ, Bertino E, Lee HC, Profit J (2016) Postnatal growth failure in very low birthweight infants born between 2005 and 2012. *Arch Dis Child Fetal Neonatal Ed* 101:F50-F55.
6. Gutmann L, Blumenthal D, Gutmann L, Schochet SS (1996) Acute type II myofiber atrophy in critical illness. *Neurology* 46:819-821.
7. Latronico N (2016) Critical illness polyneuropathy and myopathy 20 years later. No man's land? No, it is our land! *Intensive Care Med* 42:1790-1793.
8. Latronico N, Fenzi F, Recupero D, Guarneri B, Tomelleri G, Tonin P, De Maria G, Antonini L, Rizzuto N, Candiani A (1996) Critical illness myopathy and neuropathy. *Lancet* 347:1579-1582.

9. Mathai S, Derraik JG, Cutfield WS, Dalziel SR, Harding JE, Biggs J, Jefferies C, Hofman PL (2013) Increased adiposity in adults born preterm and their children. *PloS one* 8:e81840.
10. Poyhia TH, Nietosvaara YA, Remes VM, Kirjavainen MO, Peltonen JI, Lamminen AE (2005) MRI of rotator cuff muscle atrophy in relation to glenohumeral joint incongruence in brachial plexus birth injury. *Pediatr Radiol* 35:402-409.
11. Ranieri VM, Suter PM, Tortorella C, De Tullio R, Dayer JM, Brienza A, Bruno F, Slutsky AS (1999) Effect of mechanical ventilation on inflammatory mediators in patients with acute respiratory distress syndrome: a randomized controlled trial. *JAMA* 282:54-61.
12. Reid MB, Lannergren J, Westerblad H (2002) Respiratory and limb muscle weakness induced by tumor necrosis factor-alpha: involvement of muscle myofilaments. *Am J Respir Crit Care Med* 166:479-484.
13. Sher JH, Shafiq SA, Schutta HS (1979) Acute myopathy with selective lysis of myosin filaments. *Neurology* 29:100-106.
14. Tabarki B, Coffinieres A, Van Den Bergh P, Huault G, Landrieu P, Sebire G (2002) Critical illness neuromuscular disease: clinical, electrophysiological, and prognostic aspects. *Arch Dis Child* 86:103-107.
15. Thomas EL, Parkinson JR, Hyde MJ, Yap IK, Holmes E, Dore CJ, Bell JD, Modi N (2011) Aberrant adiposity and ectopic lipid deposition characterize the adult phenotype of the preterm infant. *Pediatr Res* 70:507-512.
16. Uthaya S, Thomas EL, Hamilton G, Dore CJ, Bell J, Modi N (2005) Altered adiposity after extremely preterm birth. *Pediatr Res* 57:211-215.
17. van Geleijn Vitringa VM, van Kooten EO, Jaspers RT, Mullender MG, van Doorn-Loogman MH, van der Sluijs JA (2009) An MRI study on the relations between muscle atrophy, shoulder function and glenohumeral deformity in shoulders of children with obstetric brachial plexus injury. *J Brachial Plex Periph Nerve Inj* 4:9.

- 325 18. Vassilakopoulos T, Petrof BJ (2004) Ventilator-induced diaphragmatic dysfunction.
326 Am J Respir Crit Care Med 169:336-341.
- 327 19. Weber-Carstens S, Deja M, Koch S, Spranger J, Bubser F, Wernecke KD, Spies CD,
328 Spuler S, Keh D (2010) Risk factors in critical illness myopathy during the early
329 course of critical illness: a prospective observational study. Crit Care 14:R119.
- 330
331

Table 1: Demographics, clinical parameters and DCSA in prematurely born and term infants

Data are presented as median (IQR) or *N* (%)

	Premature	Term	
	<i>N</i> =17	<i>N</i> =20	p-value
Male gender	11 (65)	9 (45)	0.325*
Gestation age (weeks)	25 (24-27)	40 (38 – 41)	<0.001
CGA (weeks)	42 (40 – 48)	41 (40 – 41)	0.005
Age (days)	113 (98 – 172)	6 (4 – 8)	<0.001
Birth weight (kg)	0.76 (0.64 – 0.93)	3.56 (3.15 – 3.87)	<0.001
Birth weight z-score	-0.47 (-1.15-0.23)	0.15 (-0.81 – 0.95)	0.065
Weight at MRI (kg)	2.98 (2.59 – 4.25)	3.65 (3.20 – 4.16)	0.185
Weight at MRI z-score	-1.61 (-3.01 - -1.08)	0.14 (-1.05 – 0.79)	<0.001
Days of invasive MV	39 (14 – 62)	4 (2 – 5)	<0.001
Days of PN	34 (19 – 52)	2 (1 – 4)	<0.001
Muscle relaxing agents	3 (18)	4 (20)	0.587
Postnatal steroids	5 (27)	2 (10)	0.212*
NEC	11 (65)	0 (0)	<0.001*
DCSA (mm ²)	189 (176 – 223)	302 (236 – 389)	<0.001

Mann-Whitney U test

*Chi square

IQR: interquartile range, CGA: corrected gestation age, MRI: Magnetic resonance imaging, MV: invasive mechanical ventilation, PN: parenteral nutrition, NEC: necrotising enterocolitis, DCSA: deltoid cross sectional area.

344 **FIGURE LEGENDS**

345

346

347 **Figure 1:** Method of free hand tracing of the perimeter of the DCSA

348 **Figure 2:** Regression analysis of DCSA with weight z-score. The regression line and 95%

349 confidence intervals are presented.

350 ▲ male infants

351 ○ female infants

352

353